



Michael Mahany. *Black Bear at Wibel Mining Camp*. Photograph. Anchorage, Alaska.

*The pathology and current concepts of classification of testicular germ cell tumors are reviewed.*

# Pathology of Germ Cell Tumors of the Testis

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**Background:** An increasing incidence of testis tumors has been noted over the second half of the 20th century. Congenital malformation of the male genitalia, prenatal risk factors, nonspecific and specific exposures in adulthood, and male infertility have all been associated with the etiology of germ cell tumors.

**Methods:** The histologic classification, pathology, and current concepts of testicular germ cell tumors are reviewed.

**Results:** Germ cell tumors occur at all ages. The tumors are identified as pure form (those of one histologic type) and mixed form (more than one histologic type). Over half of germ cell tumors consist of more than one cell type, requiring appropriate sampling for the correct diagnosis and correlation with the serum tumor markers. Burned-out germ cell tumors may occur in patients with metastatic disease with no gross evidence of a testicular tumor.

**Conclusions:** Appropriate management of testis tumors relies on accurate pathology and classification of these tumors.

## Introduction

The majority of testis tumors originate from the germ cell, which is the principal cell type of the testis. An increasing incidence of testis tumors, particularly in men of European

origin, has been noted over the second half of the 20th century.<sup>1</sup> Germ cell tumors occur at all ages. Congenital malformation of the male genitalia, prenatal risk factors, nonspecific and specific exposures in adulthood, and male infertility have all been associated with the etiology of germ cell tumors. Over half of germ cell tumors consist of more than one cell type, requiring appropriate sampling for the correct diagnosis and correlation with the serum tumor markers (human chorionic gonadotropin [hCG] and  $\alpha$ -fetoprotein [AFP]). The most recent WHO histologic classification of testis tumors<sup>2</sup> is similar to the 1998 classification,<sup>3</sup> differing most significantly with respect to the classification of teratomas and polyembryomas (Table 1). Since mature and immature teratomas have the same genetic changes and biologic potential specific for the prepubertal and postpubertal patient, teratomas are no longer subclassified as mature and immature. Polyembryoma was

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*Abbreviations used in this paper:* hCG = human chorionic gonadotropin, AFP =  $\alpha$ -fetoprotein, hPL = human placental lactogen, H&E = hematoxylin-eosin.

**Table 1. — Histologic Classification of Testis Tumors**

<b>Germ Cell Tumors</b> Intratubular germ cell neoplasia, unclassified Other types
<b>Tumors of One Histologic Type (Pure Forms)</b> Seminoma Seminoma with syncytiotrophoblastic cells Spermatocytic seminoma Embryonal carcinoma Yolk sac tumor Trophoblastic tumors Choriocarcinoma Trophoblastic neoplasms other than choriocarcinoma Monophasic choriocarcinoma Placental site trophoblastic tumor Teratoma Dermoid cyst Monodermal teratoma Teratoma with somatic type malignancies
<b>Tumors of More Than One Histologic Type (Mixed Forms)</b> Mixed embryonal carcinoma and teratoma Mixed teratoma and seminoma Choriocarcinoma and teratoma/embryonal carcinoma Others

once classified with tumors of one histologic type; however, in recognition of the different cell types present in polyembryoma, it is now classified with tumors of more than one histologic type. The tumors are staged according to the TNM classification (Table 2).<sup>4,5</sup>

## Precursor Lesions

Several terms are used to refer to intratubular malignant germ cells, including intratubular germ cell neoplasia-unclassified, carcinoma in situ, and intratubular preinvasive tumor. In the postpubertal patient, they are seen in approximately 82% of testes harboring a germ cell tumor.<sup>6</sup> However, they are rarely present in prepubertal children.<sup>7-12</sup> In testicular biopsies for infertility, intratubular malignant germ cells are seen in 0.3% to 1.8% of cases.<sup>13</sup>

There are no specific macroscopic features for these lesions. Histologically, the malignant germ cells are large with abundant pale, glycogen-rich cytoplasm. The irregularly outlined nuclei are enlarged, with one or two prominent nucleoli (Fig 1). They are located at the periphery of the seminiferous tubules between residual Sertoli cells. They often have a segmental distribution, and they can extend into the rete. At the site of invasion into the adjacent stroma, they often elicit a lymphocytic response. By immunohistology, placental alkaline phosphatase (PLAP) can be identified in up to 99% of cases with membranous and/or cytoplasmic reaction (Fig 2).<sup>6,13-15</sup> CD117a (*c-kit*)<sup>16,17</sup> shows a membranous reaction. They are strongly positive for OCT3/4 (POU5F1), a marker of pluripotent stem cells.<sup>18</sup> Intratubular malignant germ cells appear to have a similar appearance in all germ cell tumor types

with PLAP and CD117a, but other antibodies such as 43-9F<sup>19,20</sup> and TRA 1-60<sup>21,22</sup> indicate different immunophenotypes adjacent to different germ cell tumor types. Generally, the malignant germ cells are similar to the associated germ cell tumor type with respect to their DNA ploidy.<sup>23-25</sup> They do not show an increased number of isochromosome 12p [i(12p)] until they become invasive.<sup>26-28</sup>

## Tumors of One Histologic Type

### *Seminoma*

Seminomas comprise 35% to 70% of germ cell tumors, depending on the patient population (Table 3). The tumor is most commonly seen in patients between 30 and 50 years of age, and the testis is usually enlarged. The cut surface of seminomas is usually grayish-white, bulging, and glistening (Fig 3). Histologically, the tumor cells are large with abundant pale or amphophilic cytoplasm, depending on the glycogen content. The nuclei have a coarse chromatin distribution with prominent nucleoli. These cells occur in sheets, lobules, or columns. The supporting stroma shows varying amounts of lymphocytic infiltrate or granulomatous reaction (Figs 4-6). Both are prominent in approximately 20% of cases. This reaction apparently represents a host response. Immunohistochemically, the tumor cells react with antibodies to PLAP and CD117a (Table 4).<sup>6,29,30</sup> OCT4, a marker of pluripotent cells, is demonstrable in the nucleus of seminoma cells.<sup>18,31</sup> Other markers include VASA<sup>32</sup> and CD143.<sup>33</sup> The cytokeratins are occasionally positive.<sup>34</sup>

### *Variants of Seminoma*

A number of seminomas show increased mitotic activity, averaging 3 mitoses per high-power fields throughout the tumor (Fig 7). In addition, these seminomas often show more cellular atypia and pleomorphism. These tumors may express CD30, which has been interpreted by some as evidence of early differentiation toward embryonal carcinoma. This type of seminoma usually presents at a higher stage.<sup>35,36</sup> Others have questioned the significance of these findings in seminoma. When examined by immunohistochemistry, approximately 25% of seminomas contain syncytiotrophoblastic giant cells compared with 7% when examined only by hematoxylin-eosin (H&E) staining.<sup>37</sup> These are usually multinucleated cells with vacuolated cytoplasm, or they are flattened cells that are associated with vascular spaces (Fig 8). They react positive for hCG and other placental glycoproteins such as human placental lactogen (hPL) and pregnancy-specific  $\beta$ 1-glycoprotein.<sup>37,38</sup> This type of tumor is often associated with elevated serum levels of hCG.

### *Spermatocytic Seminoma*

This germ cell tumor accounts for approximately 1.2% to 4.5% of germ cell tumors.<sup>39,40</sup> Spermatocytic seminoma is

usually seen in men over 40 years of age. Approximately 5% of patients have bilateral disease.<sup>39</sup> Spermatocytic seminoma develops only in the testis, unlike other germ cell tumor types that also involve the ovary, retroperitoneum, mediastinum, and other sites. Grossly, the tumors are usually large. The cut surface is yellowish, soft, and mucoid, with cystic or spongy areas (Fig 9). Microscopically, the tumor consists of three cell types: large mononucleated or multinucleated cells with abundant cytoplasm, intermediate cells, and small cells with hyperchromatic nuclei (Fig 10). The nuclei of the large and intermediate cells have a characteristic filamentous chromatin distribution. Mitoses occur frequently. Using cytophotometry, large cells have DNA values up to 42C, whereas small cells are diploid or

near diploid and medium cells have an intermediate value. The edematous stroma is generally devoid of a lymphocytic infiltrate. Lakes of pink proteinaceous material can be seen.

The adjacent seminiferous tubules often show intratubular spermatocytic seminoma (Fig 11). The typical intratubular germ cell neoplasia unclassified is not present. About 20% of spermatocytic seminomas contain PLAP in isolated or small clusters of cells.<sup>39</sup> CD117a (*c-kit*) can also be demonstrated (I.A.S., unpublished data, 2004). AFP and hCG are not demonstrable. Most spermatocytic seminomas react diffusely with antibodies to VASA.<sup>32</sup> NY-ESO-1 has been demonstrated in spermatocytic seminoma but not in other germ cell tumor types.<sup>41</sup> Cytokeratins are usually seen in a dot-like pattern. Spermatocytic seminoma has a good

**Table 2. — TNM Classification of Tumors of the Testis**

<b>T – Primary Tumor</b>		<b>pTNM Pathological Classification (continued)</b>				
Except for pTis and pT4, where radical orchiectomy is not always necessary for classification purposes, the extent of the primary tumor is classified after radical orchiectomy; see pT. In other circumstances, TX is used if no radical orchiectomy has been performed		pN2 Metastasis with a lymph node mass more than 2 cm but not more than 5 cm in greatest dimension; or more than 5 nodes positive, none more than 5 cm; or evidence of extranodal extension of tumor				
<b>N – Regional Lymph Nodes</b>		pN3 Metastasis with a lymph node mass more than 5 cm in greatest dimension				
NX	Regional lymph nodes cannot be assessed	<b>S – Serum Tumor Markers</b>				
N0	No regional lymph node metastasis	SX Marker studies not available or not performed				
N1	Metastasis with a lymph node mass 2 cm or less in greatest dimension or multiple lymph nodes, none more than 2 cm in greatest dimension	S0 Marker study levels within normal limits				
N2	Metastasis with a lymph node mass more than 2 cm but not more than 5 cm in greatest dimension, or multiple lymph nodes, any one mass more than 2 cm but not more than 5 cm in greatest dimension	S1 LDH <1.5 × N* and hCG (mIU/mL) <5,000 and AFP (ng/mL) <1,000				
N3	Metastasis with a lymph node mass more than 5 cm in greatest dimension	S2 LDH 1.5–10 × N* or hCG (mIU/mL) 5,000–50,000 or AFP (ng/mL) 1,000–10,000				
<b>M – Distant Metastasis</b>		S3 LDH >10 × N* or hCG (mIU/mL) >50,000 or AFP (ng/mL) >10,000				
MX	Distant metastasis cannot be assessed	*N = upper limit of normal for the LDH assay.				
M0	No distant metastasis	<b>Stage Grouping</b>				
M1	Distant metastasis	Stage 0:	pTis	N0	M0	S0, SX
M1a	Non-regional lymph node(s) or lung	Stage I:	pT1–4	N0	M0	SX
M1b	Other sites	Stage IA:	pT1	N0	M0	S0
<b>pTNM Pathological Classification</b>		Stage IB:	pT2	N0	M0	S0
<b>pT – Primary Tumor</b>			pT3	N0	M0	S0
pTX	Primary tumor cannot be assessed (see T–Primary Tumor, above)		pT4	N0	M0	S0
pT0	No evidence of primary tumor (eg, histologic scar in testis)	Stage IS:	Any pT/TX	N0	M0	S1–3
pTis	Intratubular germ cell neoplasia (carcinoma in situ)	Stage II:	Any pT/TX	N1–3	M0	SX
pT1	Tumor limited to testis and epididymis without vascular/lymphatic invasion; tumor may invade tunica albuginea but not tunica vaginalis	Stage IIA:	Any pT/TX	N1	M0	S0
pT2	Tumor limited to testis and epididymis with vascular/lymphatic invasion, or tumor extending through tunica albuginea with involvement of tunica vaginalis	Stage IIB:	Any pT/TX	N1	M0	S1
pT3	Tumor invades spermatic cord with or without vascular/lymphatic invasion	Stage IIC:	Any pT/TX	N2	M0	S0
pT4	Tumor invades scrotum with or without vascular/lymphatic invasion		Any pT/TX	N2	M0	S1
<b>pN – Regional Lymph Nodes</b>		Stage III:	Any pT/TX	N3	M0	S1
pNX	Regional lymph nodes cannot be assessed	Stage IIIA:	Any pT/TX	Any N	M1, M1a	SX
pN0	No regional lymph node metastasis	Stage IIIB:	Any pT/TX	Any N	M1, M1a	S0
pN1	Metastasis with a lymph node mass 2 cm or less in greatest dimension and 5 or fewer positive nodes, none more than 2 cm in greatest dimension	Stage IIIC:	Any pT/TX	Any N	M1, M1a	S1
			Any pT/TX	N1–3	M0	S2
			Any pT/TX	Any N	M1, M1a	S2
			Any pT/TX	Any N	M1, M1a	S3
			Any pT/TX	Any N	M1b	Any S

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prognosis. Only one fully documented case of a metastasizing spermatocytic seminoma has been reported.<sup>42</sup>

### Variants of Spermatocytic Seminoma

Spermatocytic seminoma is occasionally associated with a sarcoma. Macroscopically, these tumors are large with a variegated appearance. Generally, the sarcoma is undifferentiated, but in some cases it may be differentiated, eg, rhabdomyosarcoma or chondrosarcoma.<sup>39,43-46</sup> The recognizable foci of spermatocytic seminoma often show marked nuclear pleomorphism (Fig 12). The sarcoma metastasizes extensively.

### Embryonal Carcinoma

In pure form, this tumor comprises 3% to 4% of germ cell tumors. However, it is present in approximately 40% of

tumors of more than one histologic type. It is seen most often in men between 20 and 40 years of age and less often in adolescents (15 to 20 years of age). It has not been described in the prepubertal testes. In pure form, it is not associated with elevated serum levels of AFP. Grossly, the tumors are usually small and located close to the rete. The cut surface is grayish-white with foci of hemorrhage and necrosis. They are not encapsulated (Fig 13). Microscopically, the cells are large and embryonic in appearance and have pale, amphophilic or eosinophilic cytoplasm. The cell borders are ill-defined, and there is frequent nuclear overlap. The nuclei are vesicular, with a see-through appearance. Nucleoli are prominent and mitoses are common. The growth patterns include papillary, acinar, tubular, and solid forms (Figs 14-15). The stroma is variable and may consist of loose, immature cells. In the solid pattern, degenerating cells may mimic syncytiotrophoblastic cells (Fig 15). These findings are contrary to those in seminoma, which consists of large cells with well-defined cell borders (Fig 16). Vascular and lymphatic invasion (Fig 17) and infiltration of paratesticular tissue and the epididymis are common. The adjacent seminiferous tubules may show intratubular embryonal carci-

Table 3. — Frequency of Various Histologic Types of Germ Cell Tumors in Adults

Germ Cell Tumor	Adults (%)	Infants/Children (%)
Pure seminoma	26.9	2.5
Seminoma + SCT	8.1	
Spermatocytic seminoma	2.4	
ECA	3.1	
YST	2.4	58-82
Teratoma	2.7	14-38
Choriocarcinoma	0.1	
Intratubular malignant germ cells	0.6	
ECA + YST + teratoma + SCT	14.3	
ECA + YST + teratoma + seminoma + SCT	7.4	
ECA + YST + teratoma	4.7	
YST + teratoma	2.5	0.85
ECA + teratoma + teratocarcinoma	1.4	
Teratoma + seminoma		0.45
Other combinations	24.0	

ECA = embryonal carcinoma  
YST = yolk sac tumor  
SCT = syncytiotrophoblasts

Adapted from Mostofi FK, Sesterhenn IA, Davis CJ Jr. Immunopathology of germ cell tumors of the testis. *Semin Diagn Pathol.* 1987;4:320-341. Copyright 1987, with permission from Elsevier.

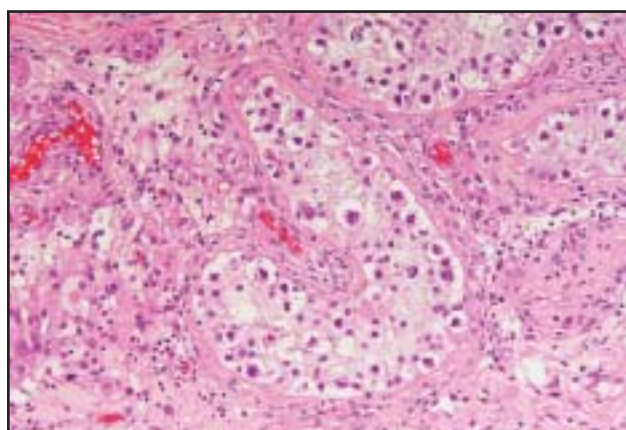


Fig 1. — Intratubular malignant germ cells (H&E, original magnification  $\times 160$ ).

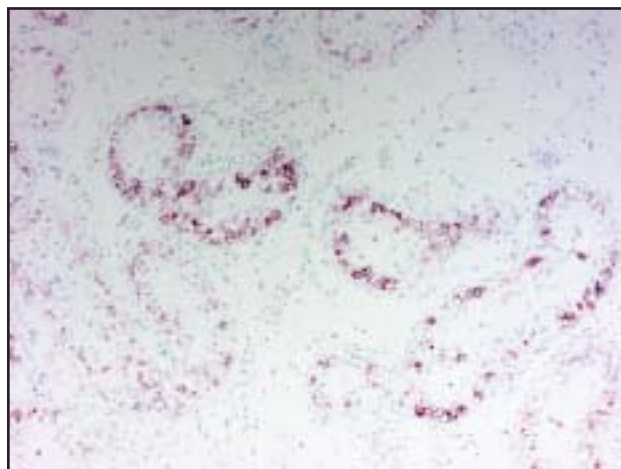


Fig 2. — Intratubular malignant germ cells with membranous and cytoplasmic staining (anti-PLAP, original magnification  $\times 80$ ).



Fig 3. — Seminoma with homogeneous pale cut surface.

noma with extensive degenerative change and calcification (Fig 18). AFP is seen in approximately 13% of cases in isolated or small clusters of cells that appear to be insufficient in number to cause a measurable elevation of serum AFP.<sup>47,48</sup> hPL can also be found.<sup>37,48</sup> OCT4 has been demonstrated in most embryonal carcinomas consistent with its pluripotentiality.<sup>18,31</sup> The presence of CD30 is common<sup>49</sup> and cytokeratins are often demonstrable, but vimentin and epithelial membrane antigen (EMA) are not. Syncytiotrophoblastic cells producing hCG can be found scattered throughout the tumor. These cells may also be positive for pregnancy-specific  $\beta$ 1-glycoprotein.<sup>37</sup> Patients with pure embryonal carcinoma or embryonal carcinoma exceeding 40% in a mixed germ cell tumor with vascular and/or lymphatic invasion often present in advanced stage.<sup>50-52</sup>

### Yolk Sac Tumor

Yolk sac tumor is the most common germ cell tumor in infants and children, accounting for approximately 65% of germ cell tumors.<sup>53</sup> It is seen in about 2.4% of adult patients, but in tumors of more than one histologic type, it

is seen in 42% of cases. On gross examination, the tumor is soft, homogenous, grayish-yellow, and not encapsulated. Microscopically, the yolk sac tumor has at least 10 different patterns, which may explain the difficulties recognizing yolk sac tumor elements in the mixed germ cell tumor. The common pattern is the reticular or microcystic pattern (Fig 19). The cells are small, ranging from cuboidal to flattened endothelial in appearance. The nuclei are of variable sizes. Mitoses are frequent, and hyaline globules are common. The solid pattern consists of cells that are smaller than seminoma cells (Fig 20). They form sheets or nodular aggregates with occasional cystic structures. If these cells are eosinophilic and resemble hepatocytes, they are of the hepatoid pattern, which is invariably positive for AFP. A glandular-alveolar pattern is not uncommon. The presence of immature glands resembling teratoma but without other teratomatous components is interpreted as the enteric pattern. The endodermal sinus pattern is characterized by papillary structures with a central fibrovascular core covered by a layer of cuboidal tumor cells. These structures are known as Schiller-Duval bodies<sup>54</sup> (Fig 21). The polyvesicular vitelline pattern consists of cysts of varying sizes separated by edematous, cellular, or fibrous stroma. In

Table 4. — Immunohistochemistry of Germ Cell Tumors

Germ Cell Tumors	Antibodies
Intratubular malignant germ cell (intratubular germ cell neoplasia, unclassified)	PLAP, CD117a, OCT4, 43-9F, TRA1-60
Seminoma	PLAP, CD117a, OCT4, VASA, cytokeratins
Spermatocytic seminoma	VASA, NY-ESO-1, cytokeratins (PLAP,* CD117a*)
Embryonal carcinoma	AFP,* OCT4, CD30, cytokeratins, PLAP, hPL
Yolk sac tumor	AFP, PLAP, cytokeratins, AAT, albumin, ferritin
Trophoblastic tumors	hCG, hPL, SP1, PLAP, Mel-CAM, HLA-G, cytokeratins
Teratoma	AFP,** PLAP, and markers specific for the different tissue types

\* rare cells  
 \*\* intestinal-like glands and hepatoid cells

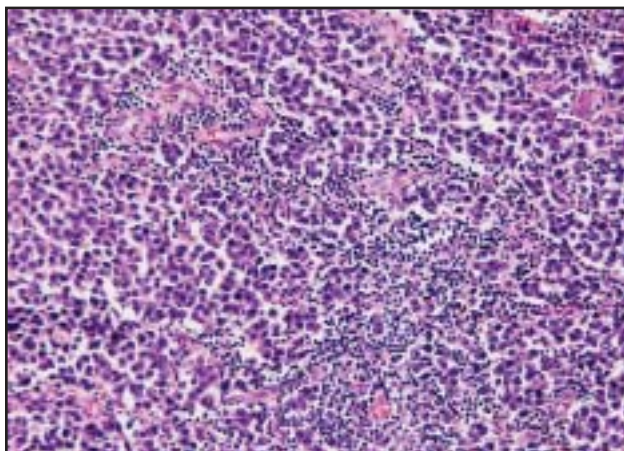


Fig 4. — Seminoma (H&E, original magnification  $\times 80$ ).

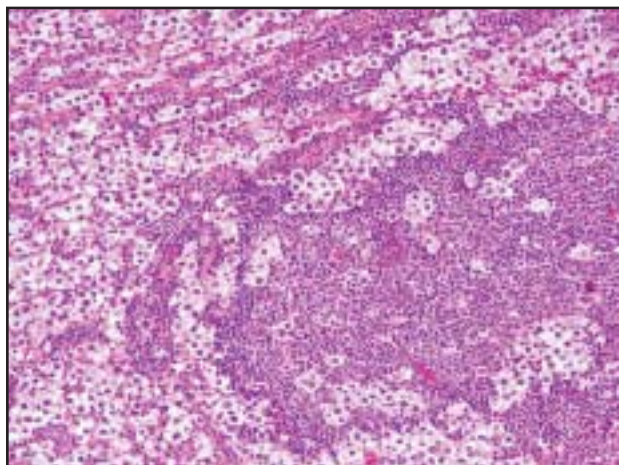


Fig 5. — Seminoma with marked lymphocytic infiltrate (H&E, original magnification  $\times 80$ ).

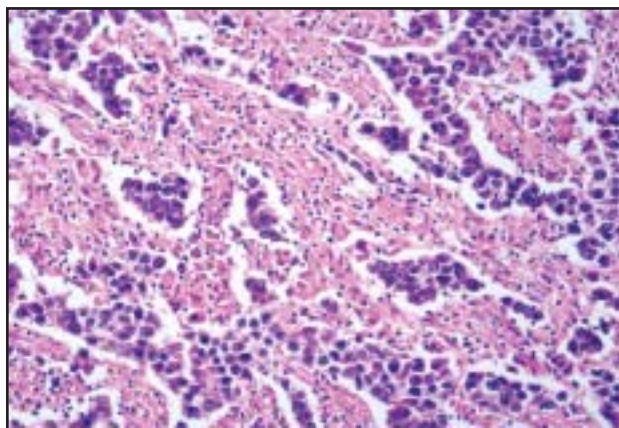


Fig 6. — Seminoma with granulomatous stroma (H&E, original magnification  $\times 100$ ).



the myxomatous pattern, the myxoid stroma predominates, with scarce epithelial elements. Immunohistochemically, AFP is focally demonstrable in approximately 92% of yolk sac tumors (Fig 22). The tumor is also positive for low-molecular-weight cytokeratin. Alpha-1 antitrypsin, albumen, ferritin, and others can be also identified.<sup>47</sup> The prognosis is similar in both children and adults.

### **Trophoblastic Tumors**

These are the rarest (>1%) of germ cell tumors. Patients often present with disseminated disease. In addition to choriocarcinoma, the monophasic choriocarcinoma and placental site trophoblastic tumors are recognized. Microscopically, the tumors are small and hemorrhagic, resembling an infarct. Peripherally, a small rim of grayish-white tissue may be seen. Histologically, the tumor consists of syncytiotrophoblastic, cytotrophoblastic, and intermediate trophoblastic cells. The syncytiotrophoblastic cells often cover nodules of cytotrophoblastic cells, forming the advancing edge (Fig 23). Hemorrhage is extensive. The syncytiotrophoblastic cells are large, with eosinophilic cytoplasm. The cytoplasm may be vacuolated, sometimes

containing erythrocytes or pale-staining eosinophilic material. They are often multinucleated but may be mononuclear. The nuclei are large and hyperchromatic. The cytotrophoblastic cells have pale-staining cytoplasm and distinct cell borders. The nuclei have 1 or 2 nucleoli. The intermediate trophoblastic cells are mononuclear cells with abundant pink cytoplasm. The syncytiotrophoblastic cells are usually positive for hCG and may have other placental glycoprotein such as human placental glycogen and pregnancy-specific  $\beta$ 1-glycoprotein. The intermediate trophoblastic cells are positive for hPL and hCG and may contain Mel-CAM and HLA-G.<sup>55</sup> PLAP and cytokeratins may be seen in all of the cells. Monophasic choriocarcinomas consisting only of cytotrophoblastic cells or intermediate trophoblasts have been reported (Fig 24).<sup>56</sup> A cystic trophoblastic tumor variant has been described following chemotherapy and occasionally in primary tumors.<sup>57</sup>

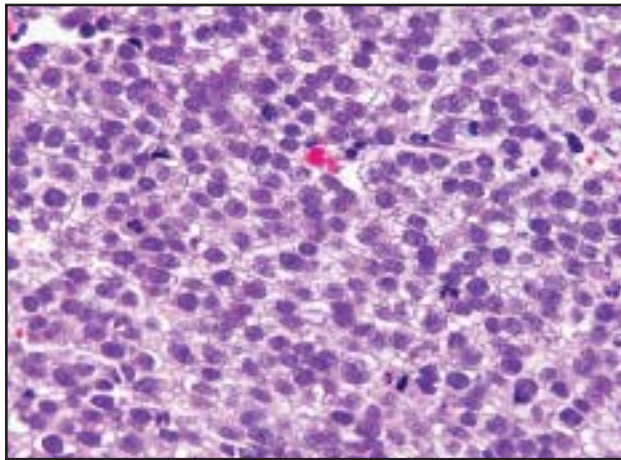


Fig 7. — Seminoma with high mitotic index; note 3 mitotic figures (H&E, original magnification  $\times 320$ ).

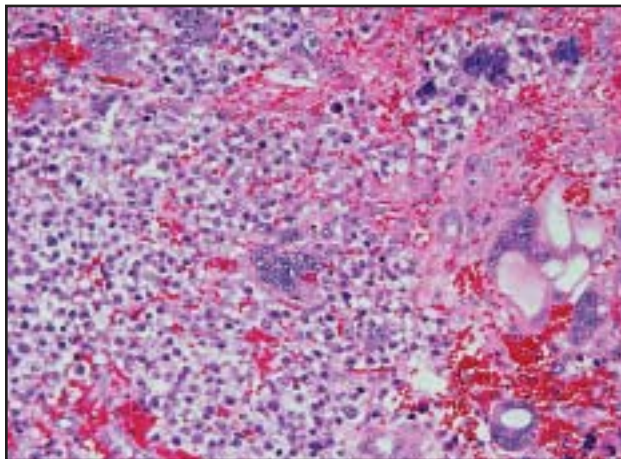


Fig 8. — Seminoma with syncytiotrophoblastic cells (H&E, original magnification  $\times 160$ ).



Fig 9. — Spermatocytic seminoma; note cystic and mucoid area.

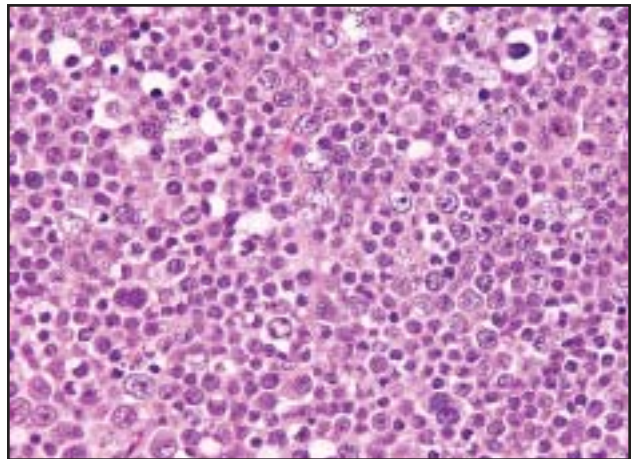


Fig 10. — Spermatocytic seminoma; note three different cell types (H&E, original magnification  $\times 320$ ).



## Teratoma

In infants and prepubertal children, teratomas constitute about 35% of germ cell tumors. They are diploid, lack chromosomal imbalances, do not show  $i(12p)$ , and are benign.<sup>58-60</sup> Since genetically mature and immature teratomas in prepubertal children are identical and their clin-

ical course is the same, separation into mature and immature teratoma is not required. In adults, they are found in 2.7% to 7% in pure form<sup>37,61</sup> but in 47% to 50% of mixed germ cell tumors.<sup>37,62,63</sup> In postpubertal patients, teratomas are hypotriploid and show chromosomal imbalances, including a gain of  $i(12p)$ .<sup>64,65</sup> They are potentially

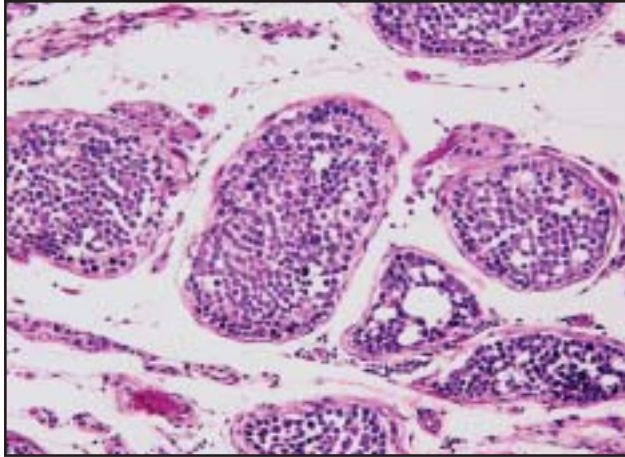


Fig 11. — Intratubular spermatocytic seminoma with 3 different cell types (H&E, original magnification  $\times 160$ ).

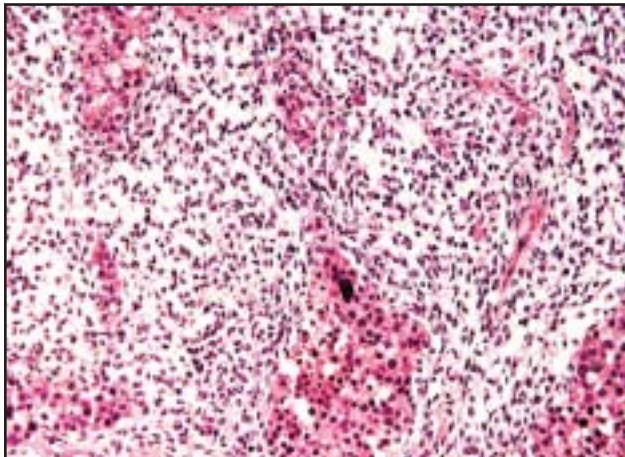


Fig 12. — Spermatocytic seminoma with sarcoma (H&E, original magnification  $\times 160$ ).



Fig 13. — Embryonal carcinoma, nodular tumor with focal hemorrhage and necrosis.

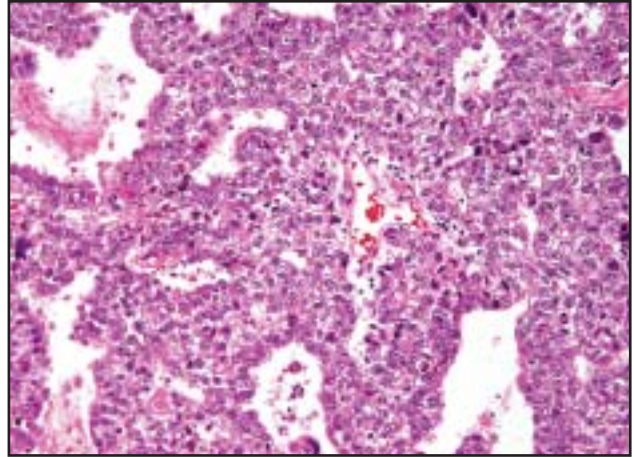


Fig 14. — Embryonal carcinoma with tubular pattern (H&E, original magnification  $\times 160$ ).

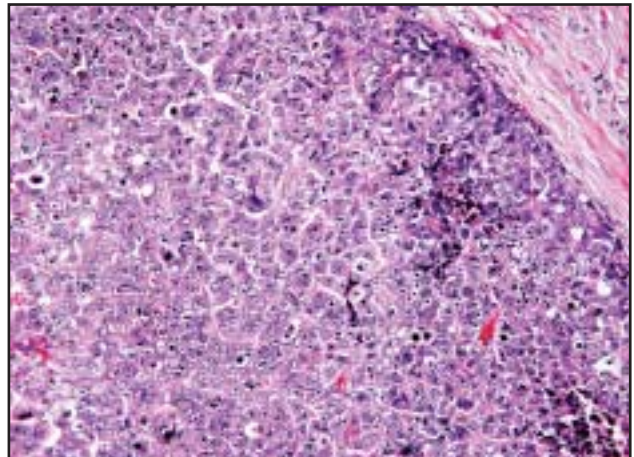


Fig 15. — Embryonal carcinoma solid pattern (H&E, original magnification  $\times 160$ ).

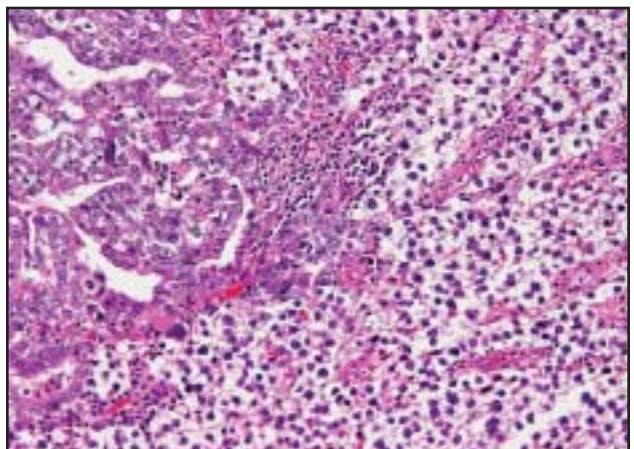


Fig 16. — Embryonal carcinoma (left) and seminoma (right) (H&E, original magnification  $\times 160$ ).



malignant since metastases can develop in up to 29% of cases. Therefore, as in the pediatric group, a distinction between mature and immature teratomas is unnecessary. However, the dermoid cyst and epidermal cyst are benign. Macroscopically, the tumors are well demarcated and show cystic areas that may contain mucoid or gelatinous material. The solid areas may consist of cartilage or bone.

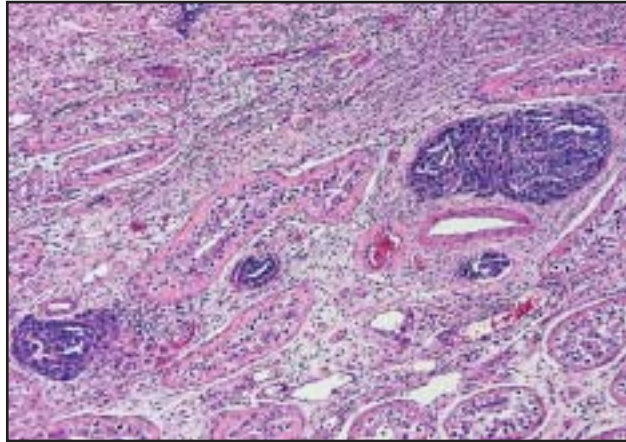


Fig 17. — Lymphovascular invasion by embryonal carcinoma (H&E, original magnification  $\times 80$ ).

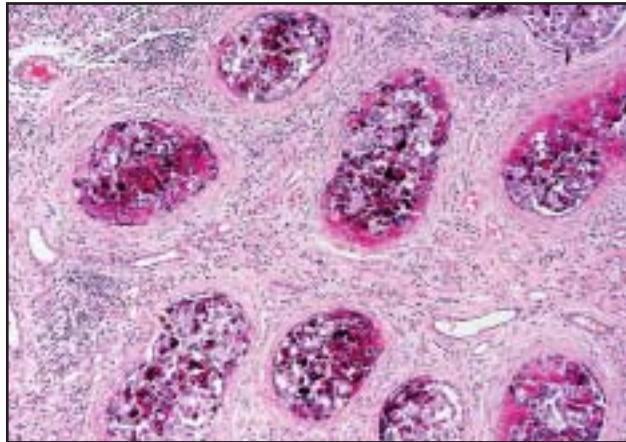


Fig 18. — Intratubular embryonal carcinoma with necrosis and calcification (H&E, original magnification  $\times 80$ ).

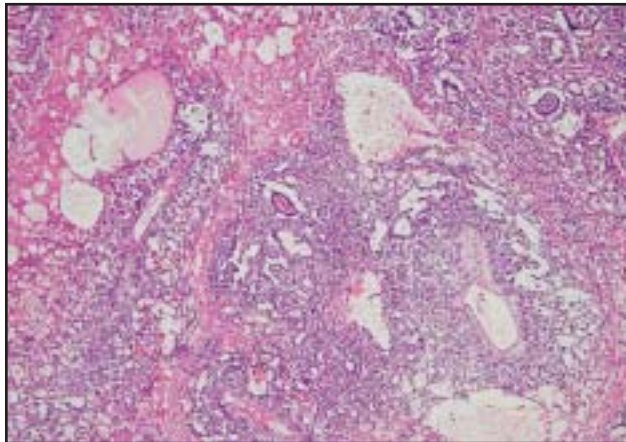


Fig 19. — Yolk sac tumor with reticular papillary growth pattern (H&E, original magnification  $\times 36$ ).

Histologically, these tumors are complex, representing the three germ cell layers: endoderm, ectoderm, and mesoderm. The tissues may be mature (Fig 25) or immature (Fig 26) in appearance. Enteric or salivary glands, respiratory epithelium, smooth and skeletal muscle, fat, cartilage, bone, glial, or neuroectodermal tissues are often haphazardly distributed. However, in infants and children, these tissue types often assume an organoid arrangement. Intratesticular and extratesticular lympho/vascular inva-

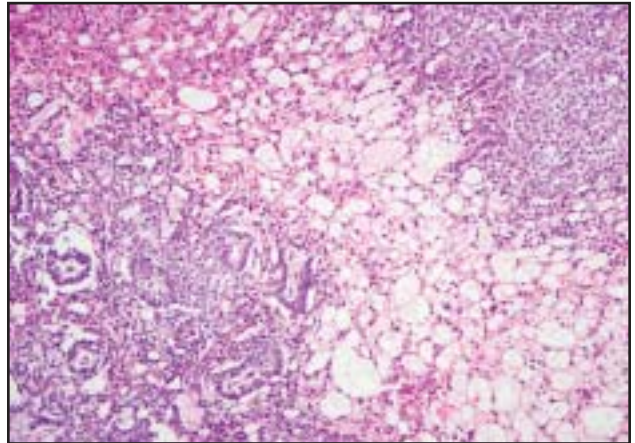


Fig 20. — Yolk sac tumor with solid reticular and papillary areas (H&E, original magnification  $\times 80$ ).

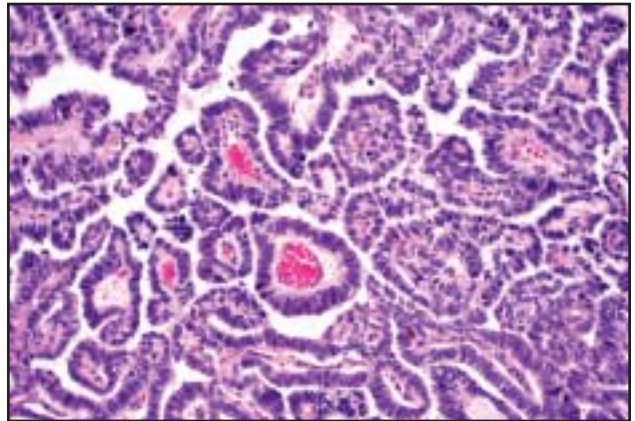


Fig 21. — Yolk sac tumor with Schiller-Duval bodies (H&E, original magnification  $\times 100$ ).

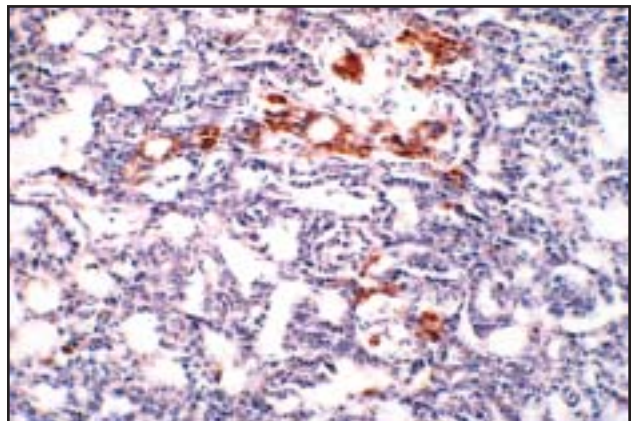


Fig 22. — Yolk sac tumor (anti-AFP, original magnification  $\times 100$ ).



sion may be seen (Fig 27), which in at least some cases explains the presence of metastases irrespective of the benign-appearing teratoma. The dermoid cyst (Fig 28) consists of a predominant cyst lined by keratinizing squamous epithelium with skin appendages. The cyst may contain a nubbin with glial tissue, bone, teeth, or other cell types. The typical intratubular malignant germ cells are not present. This tumor is benign and is rare in the testis compared with the ovary.<sup>44,66</sup>

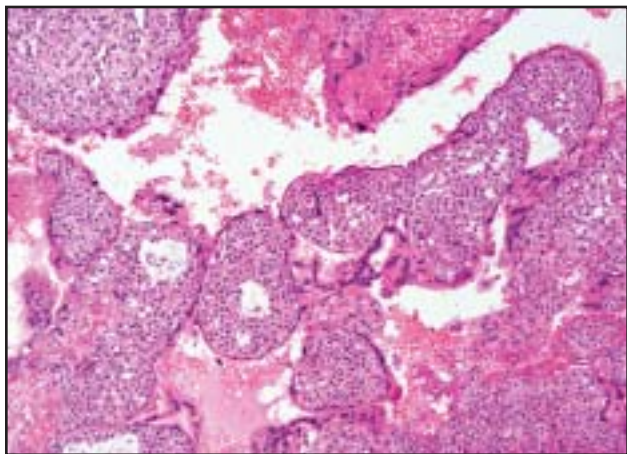


Fig 23. — Choriocarcinoma (H&E, original magnification  $\times 80$ ).

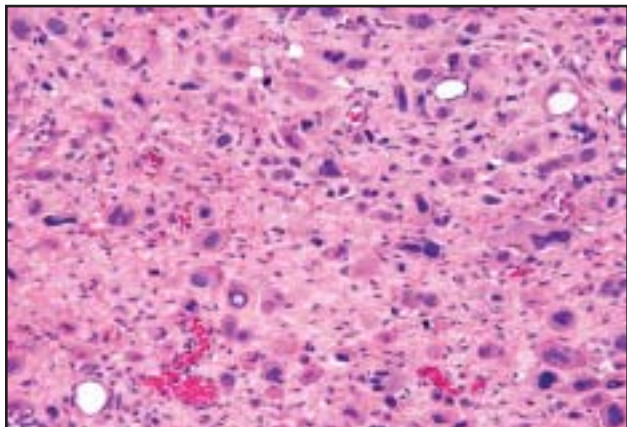


Fig 24. — Placental site trophoblastic tumor (H&E, original magnification  $\times 160$ ).

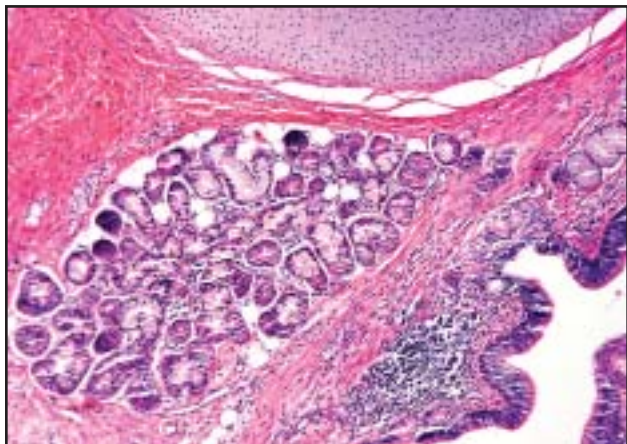


Fig 25. — Mature teratoma (H&E, original magnification  $\times 80$ ).

### Variants of Teratoma

Monodermal teratomas are rare. They consist of a single cell type (eg, cartilaginous tissue only<sup>67</sup> or peripheral neuroectodermal tumor<sup>68-71</sup>). The nature of the epidermal cyst, lined by keratinizing epithelium only, is uncertain. Some consider it a tumor-like lesion, while others favor the interpretation of a monodermal teratoma (Fig 29A-B). The latter view is supported by the presence of intratubular malignant cells in rare examples of epidermal cyst. These are benign.

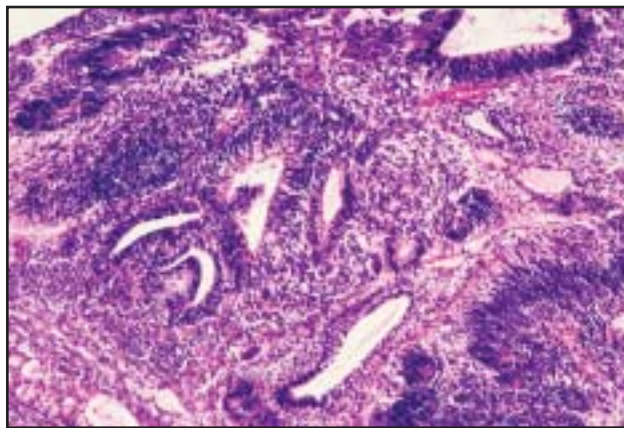


Fig 26. — Immature teratoma (H&E, original magnification  $\times 80$ ).

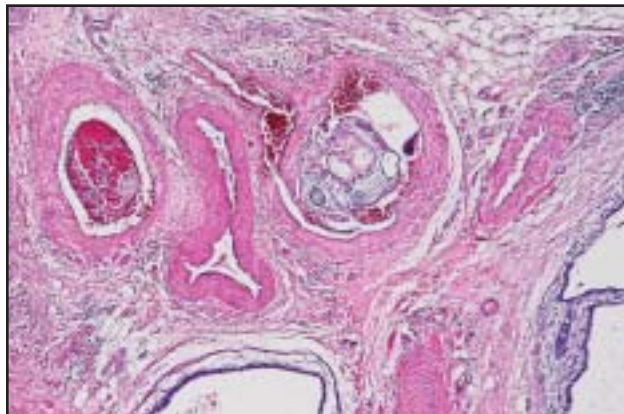


Fig 27. — Vascular invasion by teratoma and paratesticular tissue (H&E, original magnification  $\times 32$ ).

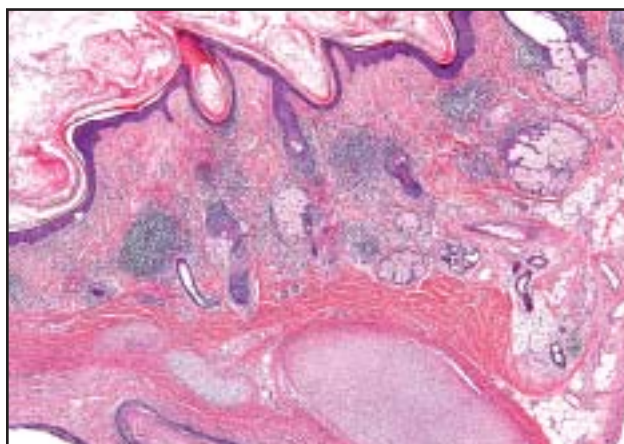


Fig 28. — Dermoid cyst (H&E, original magnification  $\times 32$ ).



Teratomas with somatic-type malignancies have malignant areas indistinguishable from those arising in other organ sites, such as rhabdomyosarcoma, adenocarcinoma (Fig 30), and nephroblastoma (Fig 31). The presence of these tumor types in the primary tumor should be included in the diagnosis since they may be the only component in metastasis, thus raising the possibility of a second primary tumor.<sup>72,73</sup> In metastases, such elements are associated with a poor prognosis.

## Tumors of More Than One Histologic Type

More than half of the germ cell tumors are a mixture of two or more of the basic germ cell tumor types, with the exception of spermatocytic seminoma. The mixed types that contain seminoma occur at a later age than those without a seminomatous component. Serum elevations of hCG indicate the presence of syncytiotrophoblastic cells either singly or as part of choriocarcinoma. Elevated serum levels of AFP are usually seen in tumors containing yolk sac elements. Teratomatous glands may also cause a serum elevation of AFP. Macroscopically, these tumors have a variegated appearance, with cystic and solid areas with or without hemorrhage or necrosis (Fig 32). Histologically, 59% of mixed germ cell tumors contain seminoma, 41% contain yolk sac tumor, and 47% contain embryonal carcinoma and teratoma. Syncytiotrophoblastic cells

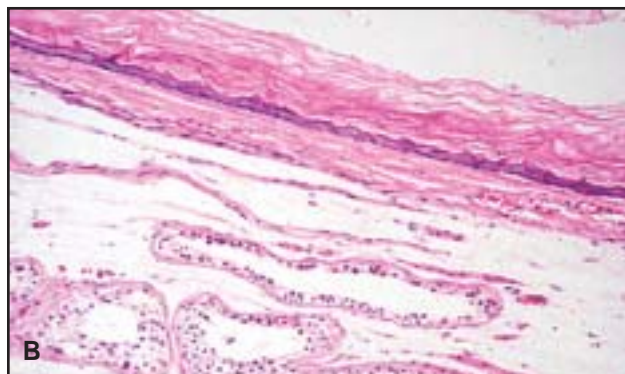


Fig 29A-B. — (A) Epidermoid cyst, note keratin lamellae with onion appearance. (B) Epidermoid cyst consisting of keratinizing squamous epithelium (H&E, original magnification  $\times 80$ ).

are present in 42%. The most common mixed germ cell tumor is the combination of teratoma, embryonal carcinoma, yolk sac tumor, and syncytiotrophoblastic cells with or without seminoma (Figs 33-35).<sup>37,52,74</sup> Polyembryoma consists of a mixture of yolk sac tumor and embryonal carcinoma forming embryoid bodies (Fig 36). They are associated with syncytiotrophoblastic cells, and a teratoma is also usually present.

The pathology report should list all of the histologic types to enable correlation with serum tumor markers. Furthermore, the percentage of each cell type should be estimated, particularly of embryonal carcinoma.<sup>51,52</sup>

## Burned-Out Germ Cell Tumor

In a number of patients with metastatic disease and no gross evidence of a testicular tumor, the only evidence of a primary tumor in that location is a homogeneous scar with remote hemorrhage.<sup>44</sup> This is commonly seen in choriocarcinomas<sup>75,76</sup> or in association with teratoma. This scar can have hematoxylin-staining bodies that contain calcium and DNA,<sup>77</sup> and it is commonly associated with intratubular malignant germ cells (Fig 37).

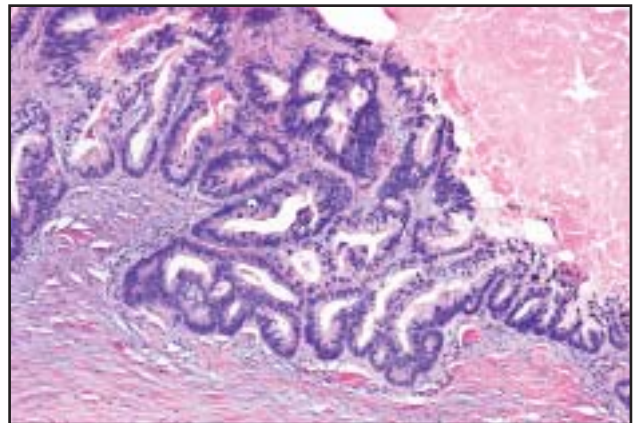


Fig 30. — Teratoma with somatic-type malignancy: adenocarcinoma (H&E, original magnification  $\times 100$ ).

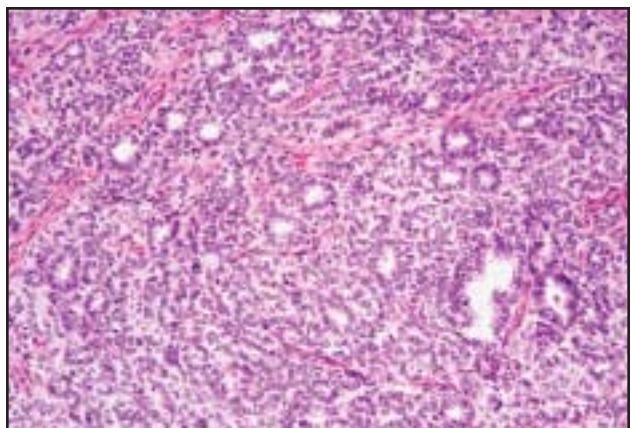


Fig 31. — Teratoma with somatic-type malignancy: nephroblastoma (H&E, original magnification  $\times 80$ ).



## Tumor Spread, Metastases, and Treatment Effects

With the exception of spermatocytic seminoma, germ cell tumor types usually develop retroperitoneal lymph node metastases. Tumors from the right testis spread to the interaortocaval, precaval, and paraaortic region with



Fig 32. — Mixed germ cell tumor. Note nodular cut surface with variegated appearance (cartilage, hemorrhage, and necrosis).

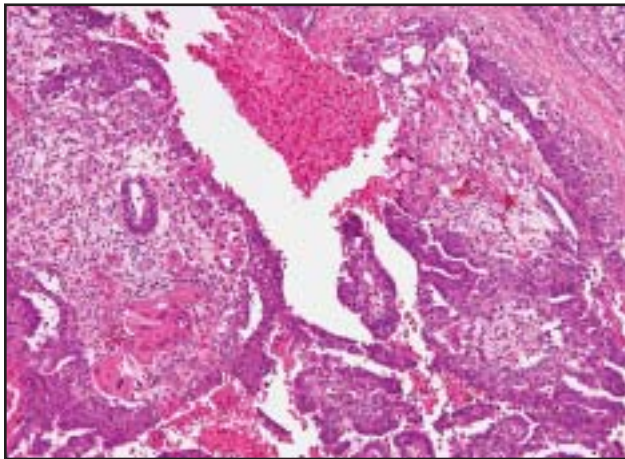


Fig 33. — Germ cell tumor consisting of teratoma, embryonal carcinoma, yolk sac tumor and syncytiotrophoblasts (H&E, original magnification  $\times 80$ ).

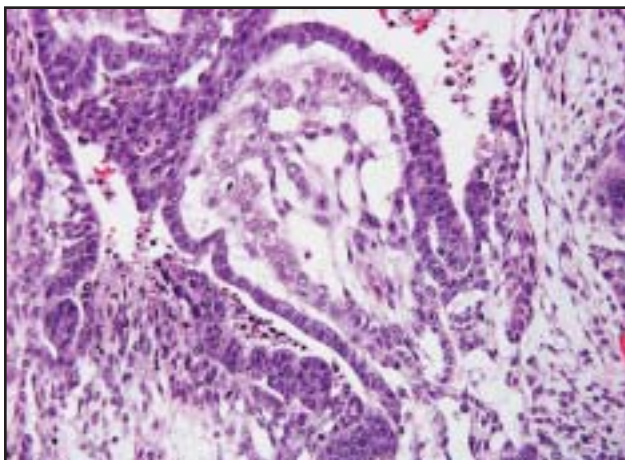


Fig 34. — Embryonal carcinoma and yolk sac tumor (H&E, original magnification  $\times 80$ ).

crossover to the left-sided lymph nodes. The left testis drains into the paraaortic and preaortic regions. Interaortocaval lymph node involvement is present in higher-stage disease.<sup>78</sup> From there, the tumors usually grow along the thoracic duct into the left supraclavicular lymph node and the subclavian vein and then show disseminated spread. Choriocarcinomas have a tendency to metastasize via the blood stream into the lung and brain. The metastases usually reflect the histology of the primary tumor. However,

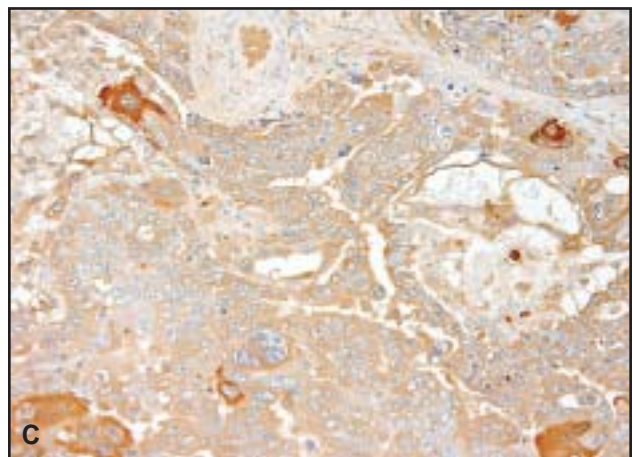
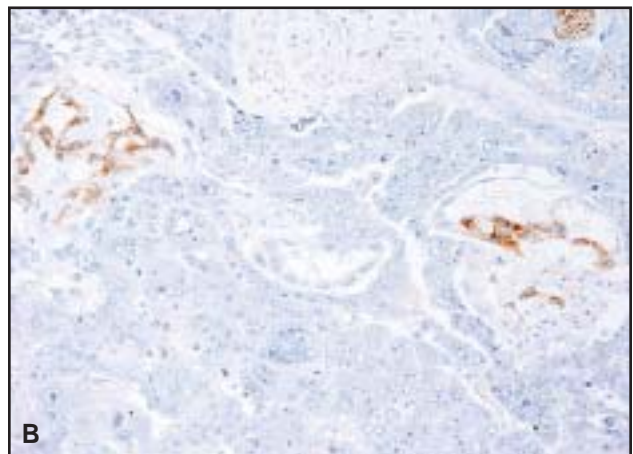
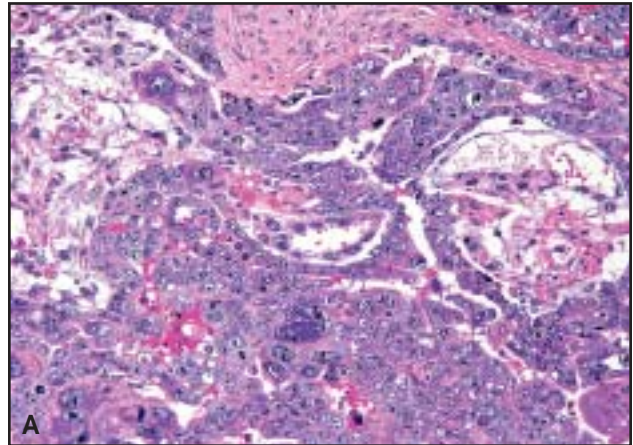


Fig 35A–C. — (A) Embryonal carcinoma, yolk sac tumor and syncytiotrophoblastic cells (H&E, original magnification  $\times 160$ ). (B) Same field (anti-AFP  $\times 160$ ). (C) Same field (anti-hCG, original magnification  $\times 160$ ).



different histologic cell types are found more often in metastases than are present in the primary tumors. This may be due to maturation of the primary germ cell type into another cell type, or it may be due to metastases from the intratubular germ cell neoplasia unclassified type that may mature at different sites to a specific germ cell tumor type. The most important predictors of metastases are the presence of vascular/lymphatic invasion in the primary tumor and the presence of embryonal carcinoma comprising over 40% of the primary tumor.<sup>51,52,79-82</sup> Chemotherapy or irradiation may result in necrosis with ghosts of tumor cells. These may be better seen in periodic acid-Schiff-stained sections. Fibrosis consists predominantly of collagen with or without calcifications and/or foreign body giant cells. Residual tumor contains teratoma in up to 40% of cases and in approximately 10% of somatic type malignancies, eg, rhabdomyosarcoma or adenocarcinoma. In rare cases, AFP may not be demonstrable in metastatic viable yolk sac tumor even though the primary tumor is positive for AFP.<sup>83</sup> Long-term follow-up is necessary since delayed metastases up to 32 years following treatment have been reported.<sup>84,85</sup>

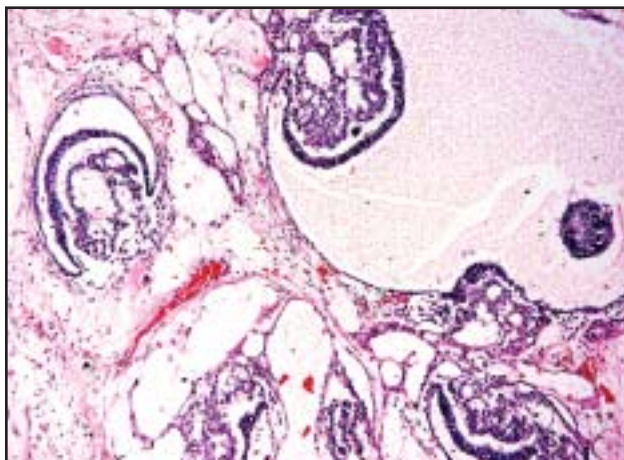


Fig 36. — Polyembryoma (H&E, original magnification  $\times 80$ ).

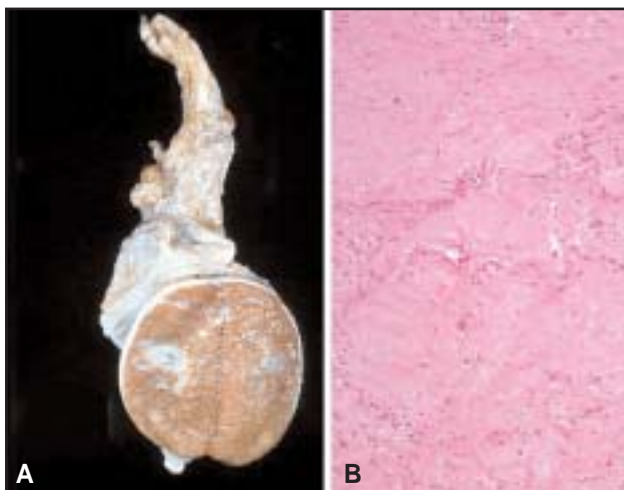


Fig 37. — (A) shows the cut surface of testis with scar; (B) shows the homogeneous acellular scar (H&E  $\times 160$ ).

*Appropriate management of testis tumors relies on accurate pathology and classification of these tumors.*

## References

1. Coleman MP, Esteve J, Damiecki P, et al. Trends in cancer incidence and mortality. *IARC Sci Publ.* 1993;121:1-806.
2. Eble JN, Sauter G, Epstein JI, Sesterhenn I. *WHO Classification of Tumours. Pathology and Genetics. Tumours of the Urinary System and Male Genital Organs.* Lyon, France: IARC Press; 2004.
3. Mostofi FK, Sesterhenn IA. *Histological Typing of Testis Tumors.* 2nd ed. World Health Organization. Springer-Verlag: Berlin Heidelberg; 1998.
4. Sobin LH, Wittekind C, eds. *TNM: Classification of Malignant Tumours.* 6th ed. Wiley & Sons: New York, NY; 2002.
5. Greene LF, Page DL, Fleming ID, et al. *AJCC Cancer Staging Manual, Sixth Edition.* New York, NY: Springer-Verlag; 2002.
6. Burke AP, Mostofi FK. Placental alkaline phosphatase immunohistochemistry of intratubular malignant germ cells and associated testicular germ cell tumors. *Hum Patol.* 1988;19:663-670.
7. Hu LM, Phillipson J, Barsky SH. Intratubular germ cell neoplasia in infantile yolk sac tumor: verification by tandem repeat sequence in situ hybridization. *Diag Mol Patol.* 1992;1:118-128.
8. Krabbe S, Skakkebaek NE, Berthelsen JG, et al. High incidence of undetected neoplasia in maldescended testes. *Lancet.* 1979;1:999-1000.
9. Parkinson MC, Swerdlow AJ, Pike MC. Carcinoma in situ in boys with cryptorchidism: when can it be detected? *Br J Urol.* 1994;73:431-435.
10. Renedo DE, Trainer TD. Intratubular germ cell neoplasia (ITGCN) with p53 and PCNA expression and adjacent mature teratoma in an infant testis: an immunohistochemical and morphologic study with a review of the literature. *Am J Surg Patol.* 1994;18:947-952.
11. Stamp IM, Barlebo H, Rix M, et al. Intratubular germ cell neoplasia in an infantile testis with immature teratoma. *Histopathology.* 1993;22:69-72.
12. Stamp IM, Jacobsen GK. Infant intratubular germ cell neoplasia. *Am J Surg Patol.* 1995;19:489.
13. Burke AP, Mostofi FK. Intratubular malignant germ cells in testicular biopsies: clinical course and identification by staining for placental alkaline phosphatase. *Mod Patol.* 1988;1:475-479.
14. Beckstead JH. Alkaline phosphatase histochemistry in human germ cell neoplasms. *Am J Surg Patol.* 1983;341-349.
15. Manivel JC, Jessurun J, Wick MR, et al. Placental alkaline phosphatase immunoreactivity in testicular germ-cell neoplasms. *Am J Surg Patol.* 1987;11:21-29.
16. Izquierdo MA, van der Valk P, van Ark-Otte J, et al. Differential expression of the c-kit proto-oncogene in germ cell tumours. *J Pathol.* 1995;177:253-258.
17. Strohmeyer T, Reese D, Press M, et al. Expression of the c-kit proto-oncogene and its ligand stem cell factor (SCF) in normal and malignant human testicular tissue. *J Urol.* 1995;153:511-515.
18. Looijenga LH, Stoop H, de Leeuw HP, et al. POU5F1 (OCT3/4) identifies cells with pluripotent potential in human germ cell tumors. *Cancer Res.* 2003;63:2244-2250.
19. Giwercman A, Lindenberg S, Kimber SJ, et al. Monoclonal antibody 43-9F as a sensitive immunohistochemical marker of carcinoma in situ of human testis. *Cancer.* 1990;65:1135-1142.
20. Heidenreich A, Sesterhenn IA, Mostofi FK, et al. Immunohistochemical expression of monoclonal antibody 43-9F in testicular germ cell tumors. *Int J Androl.* 1998;21:283-288.
21. Giwercman A, Andrews PW, Jorgensen N, et al. Immunohistochemical expression of embryonal marker TRA-1-60 in carcinoma in situ and germ cell tumors of the testis. *Cancer.* 1993;72:1308-1314.
22. Andrews PW, Banting G, Damjanov I, et al. Three monoclonal antibodies defining distinct differentiation antigens associated with different high molecular weight polypeptides on the surface of human embryonal carcinoma cells. *Hybridoma.* 1984;3:347-361.



23. de Graaff WE, Oosterhuis JW, de Jong B, et al. Ploidy of testicular carcinoma in situ. *Lab Invest.* 1992;66:166-168.
24. el-Naggar AK, Ro JY, McLemore D, et al. DNA ploidy in testicular germ cell neoplasms: histogenetic and clinical implications. *Am J Surg Pathol.* 1992;16:611-618.
25. Muller J, Skakkebaek NE. Microspectrophotometric DNA measurements of carcinoma in situ germ cells in testis. *Int J Androl.* 1981;4:211-221.
26. Looijenga LH, Rosenberg C, van Gurp RJ, et al. Comparative genomic hybridization of microdissected samples from different stages in the development of a seminoma and a non-seminoma. *J Pathol.* 2000;191:187-192.
27. Rodriguez E, Houldsworth J, Reuter VE, et al. Molecular cytogenetic analysis of i(12p)-negative human male germ cell tumors. *Genes Chromosomes Cancer.* 1993;8:230-236.
28. Rosenberg C, van Gurp RJ, Geelen E, et al. Overrepresentation of the short arm of chromosome 12 is related to invasive growth of human testicular seminomas and nonseminomas. *Oncogene.* 2000;19:5858-5862.
29. Ulbright TM, Amin MB, Young RH. *Atlas of Tumor Pathology: Tumors of the Testis, Adnexa, Spermatic Cord and Scrotum.* Washington, DC: Armed Forces Institute of Pathology; 1999.
30. Leroy X, Augusto D, Leteurtre E, et al. CD30 and CD117 (c-kit) used in combination are useful for distinguishing embryonal carcinoma from seminoma. *J Histochem Cytochem.* 2002;50:283-285.
31. Jones TD, Ulbright TM, Eble JN, et al. OCT4 staining in testicular tumors. a sensitive and specific marker for seminoma and embryonal carcinoma. *Am J Surg Pathol.* 2004;28:935-940.
32. Zeeman AM, Stoop H, Boter M, et al. VASA is a specific marker for both normal and malignant human germ cells. *Lab Invest.* 2002; 82:159-166.
33. Franke FE, Pauls K, Metzger R, et al. Angiotensin I-converting enzyme and potential substrates in human testis and testicular tumours. *APMIS.* 2003;111:234-244.
34. Chevillet JC, Rao S, Iczkowski KA, et al. Cytokeratin expression in seminoma of the human testis. *Am J Clin Pathol.* 2000;113:583-588.
35. Tickoo SK, Hutchinson B, Bacik J, et al. Testicular seminoma: a clinicopathologic and immunohistochemical study of 105 cases with special reference to seminomas with atypical features. *Int J Surg Pathol.* 2002;10:23-32.
36. Thackray AC, Crane WA. Seminoma. In: Pugh RCB, ed. *Pathology of the Testis.* Blackwell Scientific: Oxford; 1976:164-198.
37. Mostofi FK, Sesterhenn IA, Davis CJ Jr. Immunopathology of germ cell tumors of the testis. *Semin Diagn Pathol.* 1987;4:320-341.
38. Jacobsen GK, von der Maase H, Specht L. Histopathological features of stage I seminoma treated with orchidectomy only. *J Urol Pathol.* 1995;3:85-94.
39. Burke AP, Mostofi FK. Spermatocytic seminoma: a clinicopathologic study of 79 cases. *J Urol Pathol.* 1993;1:21-32.
40. Jacobsen GK, Barlebo H, Olsen J. Testicular germ cell tumors in Denmark 1976-1980: pathology of 1058 consecutive cases. *Acta Radiol Oncol.* 1984;23:293-347.
41. Satie AP, Rajpert-De Meyts E, Spagnoli GC, et al. The cancer-testis gene, NY-ESO-1, is expressed in normal fetal and adult testes and in spermatocytic seminomas and testicular carcinoma in situ. *Lab Invest.* 2002;82:775-780.
42. Matoska J, Ondrus D, Hornak M. Metastatic spermatocytic seminoma: a case report with light microscopic, ultrastructural, and immunohistochemical findings. *Cancer.* 1988;62:1197-1201.
43. Matoska J, Talerma A. Spermatocytic seminoma associated with rhabdomyosarcoma. *Am J Clin Pathol.* 1990;94:89-95.
44. Mostofi FK, Price EB. *Tumors of the Male Genital System.* Washington, DC: Armed Forces Institute of Pathology; 1973.
45. True LD, Otis CN, Delprado W, et al. Spermatocytic seminoma of testis with sarcomatous transformation: a report of five cases. *Am J Surg Pathol.* 1988;12:75-82.
46. Floyd C, Ayala AG, Logothetis CJ, et al. Spermatocytic seminoma with associated sarcoma of the testis. *Cancer.* 1988;61:409-414.
47. Jacobsen GK, Jacobsen M. Alpha-fetoprotein (AFP) and human chorionic gonadotropin (HCG) in testicular germ cell tumours: a prospective immunohistochemical study. *Acta Pathol Microbiol Immunol Scand [A].* 1983;91:165-176.
48. Jacobsen GK, Jacobsen M, Clausen PP. Distribution of tumor-associated antigens in the various histologic components of germ cell tumors of the testis. *Am J Surg Pathol.* 1981;5:257-266.
49. Pallesen G, Hamilton-Dutoit SJ. Ki-1 (CD30) antigen is regularly expressed by tumor cells of embryonal carcinoma. *Am J Pathol.* 1988;133:446-450.
50. Mostofi FK, Sesterhenn IA. Pathology of germ cell tumors of testes. *Prog Clin Biol Res.* 1985;203:1-34.
51. Moul JW, McCarthy WF, Fernandez EB, et al. Percentage of embryonal carcinoma and of vascular invasion predicts pathological stage in clinical stage I nonseminomatous testicular cancer. *Cancer Res.* 1994;54:362-364.
52. Sesterhenn IA, Weiss RB, Mostofi FK, et al. Prognosis and other clinical correlates of pathologic review in stage I and II testicular carcinoma: a report from the Testicular Cancer Intergroup Study. *J Clin Oncol.* 1992;10:69-78.
53. Kaplan GW, Cromie WC, Kelalis PP, et al. Prepubertal yolk sac testicular tumors: report of the testicular tumor registry. *J Urol.* 1988;140:1109-1112.
54. Teilum G. Endodermal sinus tumors of the ovary and testis: comparative morphogenesis of the so-called mesonephroma ovarii (schiller) and extraembryonic (yolk sac-allantoic) structures of the rat placenta. *Cancer.* 1959;12:1092-1105.
55. Singer G, Kurman RJ, McMaster MT, et al. HLA-G immunoreactivity is specific for intermediate trophoblast in gestational trophoblastic disease and can serve as a useful marker in differential diagnosis. *Am J Surg Pathol.* 2002;26:914-920.
56. Ulbright TM, Young RH, Scully RE. Trophoblastic tumors of the testis other than classic choriocarcinoma: "monophasic" choriocarcinoma and placental site trophoblastic tumor: a report of two cases. *Am J Surg Pathol.* 1997;21:282-288.
57. Cheng L, Henley JD, Cummings OW, et al. Cystic trophoblastic tumor: a favorable histologic lesion in post-chemotherapy resections of patients with testicular germ cell tumors. *Mod Pathol.* 2001;14:104A. Abstract.
58. Rushton HG, Belman AB, Sesterhenn I, et al. Testicular sparing surgery for prepubertal teratoma of the testis: a clinical and pathological study. *J Urol.* 1990;144:726-730.
59. Silver SA, Wiley JM, Perlman EJ. DNA ploidy analysis of pediatric germ cell tumors. *Mod Pathol.* 1994;7:951-956.
60. Mostert M, Rosenberg C, Stoop H, et al. Comparative genomic and in situ hybridization of germ cell tumors of the infantile testis. *Lab Invest.* 2000;80:1055-1064.
61. Friedman NB, Moore RA. Tumors of the testis: a report on 922 cases. *Milit Surgeon.* 1946;99:573-593.
62. Barsky SH. Germ cell tumors of the testis. In: *Surgical Pathology of Urology Diseases*, Javadpour N, Barsky SH, eds. Baltimore, Md: Williams and Wilkins; 1987.
63. von Hochstetter AR, Hedinger CE. The differential diagnosis of testicular germ cell tumors in theory and practice: a critical analysis of two major systems of classification and review of 389 cases. *Virchows Arch A Pathol Anat Histol.* 1982;396:247-277.
64. Mostert MC, Verkerk AJ, van de Pol M, et al. Identification of the critical region of 12p over-representation in testicular germ cell tumors of adolescents and adults. *Oncogene.* 1998;16:2617-2627.
65. Mostert MM, van de Pol M, Olde Weghuis D, et al. Comparative genomic hybridization of germ cell tumors of the adult testis: confirmation of karyotypic findings and identification of a 12p-amplicon. *Cancer Genet Cytogenet.* 1996;89:146-152.
66. Ulbright TM, Srigley JR. Dermoid cyst of the testis: a study of five postpubertal cases, including a pilomatixoma-like variant, with evidence supporting its separate classification from mature testicular teratoma. *Am J Surg Pathol.* 2001;25:788-793.
67. Singh N, Cumming J, Theaker JM. Pure cartilaginous teratoma differentiated of the testis. *Histopathology.* 1997;30:373-374.
68. Aguirre P, Scully RE. Primitive neuroectodermal tumor of the testis: report of a case. *Arch Pathol Lab Med.* 1983;107:643-645.
69. Nistal M, Paniagua R. Primary neuroectodermal tumour of the

- testis. *Histopathology*. 1985;9:1351-1359.
70. Nocks BN, Dann JA. Primitive neuroectodermal tumor (immature teratoma) of testis. *Urology*. 1983;22:543-544.
  71. Young RH, Scully RE. *Testicular Tumors*. Chicago, Ill: ASCP Press; 1990.
  72. Motzer RJ, Amsterdam A, Prieto V, et al. Teratoma with malignant transformation: diverse malignant histologies arising in men with germ cell tumors. *J Urol*. 1998;159:133-138.
  73. Ahmed T, Bosl GJ, Hajdu SI. Teratoma with malignant transformation in germ cell tumors in men. *Cancer*. 1985;56:860-863.
  74. Mostofi FK. Pathology of germ cell tumors of testis: a progress report. *Cancer*. 1980;45:1735-1754.
  75. Lopez JI, Angulo JC. Burned-out tumour of the testis presenting as retroperitoneal choriocarcinoma. *Int Urol Nephrol*. 1994;26:549-553.
  76. Rottinto A, Debellis H. Extragenital chorioma: its relation to teratoid vestiges in the testicles. *Arch Pathol*. 1944;37:78-80.
  77. Azzopardi JG, Mostofi FK, Theiss EA. Lesions of testes observed in certain patients with widespread choriocarcinoma and related tumors: the significance and genesis of hematoxylin-staining bodies in human testis. *Am J Pathol*. 1961;38:207-225.
  78. Donohue JP, Zachary JM, Maynard BR. Distribution of nodal metastases in nonseminomatous testis cancer. *J Urol*. 1982;128:315-320.
  79. Borge N, Fossa SD. Late relapses of testicular cancer: a review. *Cancer J*. 1990;3:53-55.
  80. Roth BJ, Greist A, Kubilis PS, et al. Cisplatin-based combination chemotherapy for disseminated germ cell tumors: long-term follow-up. *J Clin Oncol*. 1988;6:1239-1247.
  81. Freedman LS, Parkinson MC, Jones WG, et al. Histopathology in the prediction of relapse of patients with stage I testicular teratoma treated by orchidectomy alone. *Lancet*. 1987;2:294-298.
  82. Fung CY, Kalish LA, Brodsky GL, et al. Stage I nonseminomatous germ cell testicular tumor: prediction of metastatic potential by primary histopathology. *J Clin Oncol*. 1988;6:1467-1473.
  83. Mostofi FK. Histological change ostensibly induced by therapy in the metastasis of germ cell tumors of testis. *Prog Clin Biol Res*. 1985;203:47-60.
  84. Blanke CD, Delgalvis SC, Nichols GR. Late recurrence of seminoma. *South Med J*. 1997;90:653-655.
  85. Baniel J, Foster RS, Gonin R, et al. Late relapse of testicular cancer. *J Clin Oncol*. 1995;13:1170-1176.